

Hypersensitivity Pneumonitis

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KEYWORDS

- Extrinsic allergic alveolitis • Farmer's lung • Bird fancier's disease • HRCT
- Bronchoalveolar lavage • Prognosis

KEY POINTS

- Clinical manifestations of hypersensitivity pneumonitis may closely mimic other interstitial lung diseases, and the disease onset is usually insidious.
- High-resolution computed tomography and bronchoalveolar lavage are the sensitive and characteristic diagnostic tests for hypersensitivity pneumonitis.
- The relevant antigen to hypersensitivity pneumonitis cannot be identified in up to 20% to 30% of patients.
- Clinicians should be aware that hypersensitivity pneumonitis must be considered in all cases of interstitial lung disease, and a detailed environmental exposure history is mandatory.

INTRODUCTION

Hypersensitivity pneumonitis (HP), synonymous with extrinsic allergic alveolitis (EAA), is a complex syndrome resulting from repeated exposure to a variety of antigenic particles found in the environment.¹ Because the resulting inflammatory response involves not only the alveoli but the terminal bronchioli and the interstitium, the term HP may be more correct.

The prevalence of HP is difficult to determine, because the disease is often unrecognized or misdiagnosed. The estimated prevalence of farmer's lung ranges from 1% to 19% of exposed farmers,^{2–4} the prevalence of pigeon breeder's lung is from 6% to 20% of exposed individuals,⁵ and the prevalence of budgerigar's lung is from 1% to 8% of

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exposed individuals.⁶ The disease may also arise in children. Clinical behavior in children is similar to adult cases.^{7,8}

The clinical manifestations have regional characteristics. Farmer's lung and pigeon breeder's lung are more common in cold and wet regions, mainly in Europe, whereas summer-type HP is limited to Japan.

A wide variety of particles sized less than 5 μm can reach the alveoli and may be the pathogens of HP. The causative particles include fungal (ie, *Aspergillus* and *Penicillium* species), bacterial, protozoal, animal (mostly bird) and insect proteins, and low-molecular-weight chemical compounds (ie, isocyanates, zinc, inks, and dyes) (Table 1).⁹

More recent studies have suggested that mist from a domestic ultrasonic humidifier,¹⁰ steam iron,¹¹ dry sausage dust,^{12,13} wind instruments including saxophone^{14,15} and trombone,¹⁶ colistin,¹⁷ catechin (green tea extract),¹⁸ and methylmethacrylate (in dental technicians)¹⁹ can be the cause of HP. Feather duvet lung has been reported as a rare subgroup of bird fancier's lung.^{20,21}

HP may present as acute, subacute, or chronic clinical forms, but these forms frequently overlap. The clinical presentation of HP is influenced by several factors including the nature and the amount of inhaled antigen, the intensity and frequency of exposure, and the host immune response, which is likely determined by a genetic background (Table 2).¹

It is not known why HP develops only in a minority of exposed individuals, or why some cases of chronic HP show progression without further antigen exposure.

CLINICAL FEATURES

The spectrum of clinical features varies and has been conventionally classified into acute, subacute, and chronic forms. The interval between sensitization by antigen inhalation and the symptomatic onset of HP is unknown. It seems to be variable and may range from several months to several years after the antigen exposure.

Acute Form

Acute HP is characterized by an influenzalike syndrome (fever, chills, malaise, myalgia, headache) and respiratory symptoms (dry cough, dyspnea, tachypnea, chest tightness). However, respiratory symptoms in acute HP are sometimes absent. The disease onset is abrupt and usually occurs 4 to 12 hours after antigen exposure. In general, acute HP is nonprogressive and spontaneously improves within a few days after antigen avoidance.¹ The disease often recurs after the reexposure of antigen. The clinical examination shows bibasilar crackles and occasional cyanosis, whereas finger clubbing is rare. Patients with recurrent acute farmer's lung may sometimes develop an obstructive lung disease with centrilobular emphysema instead of fibrosis.²²

Subacute Form

Subacute HP may be associated with repeated low-level exposure to inhaled antigens.²³ After recurrent acute episodes, this form may also become chronic, resulting in fibrosis. It is characterized by an insidious onset of dyspnea, fatigue, and cough. Because the respiratory symptoms are usually mild or absent in subacute HP, infectious pneumonia or noninfectious interstitial lung disease (ILD) is the important differential diagnosis.

Chronic Form

Chronic HP may result from continuous, low-level exposure to inhaled antigens.²³ Bird antigen exposure is the most common in this form of disease. The onset of chronic HP is insidious with slowly increasing dyspnea, dry cough, fatigue, and weight loss. Digital

Table 1 Environmental exposure and antigens in various types of HP		
Disease	Exposure	Antigen
Microorganisms		
Farmer's lung	Moldy hay, grain	<i>Saccharospora rectivirgula</i> , <i>Thermoactinomyces vulgaris</i> , <i>Aspergillus</i> spp
Humidifier lung; air conditioner lung	Contaminated humidifiers and air conditioners	Amoebae, nematodes, yeasts, bacteria
Misting fountain HP	Contaminated water	Bacteria, molds, yeasts
Steam iron HP	Contaminated water reservoir	<i>Sphingobacterium spiritivorum</i>
Suberosis	Moldy cork	<i>Penicillium</i> spp
Sequoiosis	Moldy redwood dust	<i>Graphium</i> spp, <i>Pullularia</i> spp, <i>Trichoderma</i> spp
Woodworker's lung	Contaminated wood pulp or dust	<i>Alternaria</i> spp
Wood-trimmer's lung	Contaminated wood trimmings	<i>Rhizopus</i> spp, <i>Mucor</i> spp
Maple-bark stripper's lung	Contaminated maple logs	<i>Cryptostroma corticale</i>
Domestic allergic alveolitis	Decayed wood	Molds
Sauna-taker's lung	Contaminated sauna water	<i>Aureobasidium</i> spp
Basement lung	Contaminated basements	<i>Cephalosporium</i> spp, <i>Penicillium</i> spp
Hot-tub lung	Mold on ceiling, tub water	<i>Mycobacterium avium</i> complex
Swimming pool lung	Mist from pool water, sprays and fountains	<i>M avium</i> complex
Thatched roof lung	Dried grasses and leaves	<i>Saccharomonospora viridis</i> , <i>T vulgaris</i> , <i>Aspergillus</i> spp
Bagassosis	Moldy pressed sugar cane (bagasse)	<i>Thermoactinomyces sacchari</i> , <i>T vulgaris</i>
Mushroom-worker's lung	Moldy compost and mushrooms	<i>S rectivirgula</i> , <i>T vulgaris</i> , <i>Aspergillus</i> spp
Malt-worker's lung	Contaminated barley	<i>Aspergillus clavatus</i>
Cheese-washer's lung	Moldy cheese or cheese casings	<i>Penicillium casei</i>
Dry sausage worker's lung	Moldy sausage dust	<i>Penicillium</i> spp
Paprika slicer's lung	Moldy paprika pods	<i>Mucor stolonifer</i>
Compost lung	Compost	<i>Aspergillus</i> spp, <i>T vulgaris</i>
Wine-maker's lung	Mold on grapes	<i>Botrytis cinerea</i>
Tobacco-grower's lung	Mold on tobacco	<i>Aspergillus</i> spp
Potato-riddler's lung	Moldy hay around potatoes	Thermophilic actinomycetes, <i>Aspergillus</i> spp
Summer-type HP	Contaminated houses	<i>Trychosporon cutaneum</i>
Detergent lung, washing powder lung	Detergents (during processing or use)	<i>Bacillus subtilis</i> enzymes

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Table 1 (continued)		
Disease	Exposure	Antigen
Machine-operator's lung	Contaminated metalworking fluid	<i>Pseudomonas</i> spp, nontuberculous mycobacteria <i>Aspergillus fumigatus</i>
Stipatosis	Esparto dust	Thermophilic actinomycetes
Peat moss HP	Contaminated peat moss	<i>Monocillium</i> spp; <i>Penicillium citreonigum</i>
Wind-instrument lung	Contaminated saxophones, trombone	Molds, bacteria
Chiropodist's lung	Foot skin and nail dust	Fungi
Animal proteins		
Bird fancier's lung; pigeon breeder's lung	Parakeets, budgerigars, pigeons, parrots, cockatiels, chickens, turkeys, geese, ducks, lovebirds	Proteins in avian droppings, in serum, and on feathers
Feather duvet lung	Feather beds, pillows, duvets	Avian proteins
Pituitary snuff-taker's lung	Bovine and porcine pituitary powder	Pituitary proteins
Furrier's lung	Animal pelts	Animal fur dust
Animal handler's lung, laboratory worker's lung	Rats, gerbils	Proteins from urine, serum, pelts
Pearl oyster shell HP	Dust of shells	Pearl oyster proteins
Mollusk shell HP	Sea snail shell dust	Sea snail shell protein
Silk production HP	Dust from silkworm larvae and cocoons	Silkworm proteins
Miller's lung	Contaminated grain	<i>Sitophilus granarius</i> (wheat weevil)
Chemicals		
Chemical worker's lung	Polyurethane foams, spray paints, elastomers, glues	Diisocyanates, trimellitic anhydride
Epoxy resin lung	Heated epoxy resin	Phthalic anhydride
Unknown		
Mummy-handler's lung	Cloth wrappings of mummies	—
Coffee-worker's lung	Coffee-bean dust	—
Tap water lung	Contaminated tap water	—
Tea-grower's lung	Tea plants	—

Adapted from Costabel U, Bonella F, Guzman J. Chronic hypersensitivity pneumonitis. Clin Chest Med 2012; 33:151–63.

clubbing may be present in 20% to 50% of patients^{1,23} and predicts clinical deterioration.²⁴ Chronic HP often develops progressive fibrosis with cor pulmonale and mimics idiopathic pulmonary fibrosis (IPF) or fibrotic nonspecific interstitial pneumonia (NSIP) in the advanced stage.²⁵ This form of disease, therefore, often leads the

Table 2
Symptoms and signs in 116 patients with HP

Feature	Frequency (%)
Dyspnea	98
Cough	91
Chills	34
Fever	19
Chest tightness	35
Weight loss	42
Body aches	24
Wheezing	31
Inspiratory crackles	87
Cyanosis	32
Clubbing	21

Adapted from Costabel U, Bonella F, Guzman J. Chronic hypersensitivity pneumonitis. Clin Chest Med 2012; 33:151–63.

physician to mistake the disease for other chronic ILDs. The auscultatory findings include bibasilar crackles and characteristically inspiratory squeaks resulting from the coexisting bronchiolitis.

Acute Exacerbation

Acute exacerbation of chronic HP is an emerging concept showing an accelerated respiratory deterioration with the presence of new bilateral ground-glass opacities on high-resolution computed tomography (HRCT).^{26,27} The pathogenesis of acute exacerbations in chronic HP is unknown.

It is likely to occur without further exposure to the inhaled antigens. Low total lung capacity (TLC) and diffusing capacity of the lung for carbon monoxide (D_{LCO}), fewer lymphocytes and increased neutrophils in bronchoalveolar lavage (BAL) fluids, and a UIP-like pattern in histology at the time of diagnosis seem to be the risk factors for acute exacerbation.²⁶ Pathologic findings include organizing pneumonia (OP) or diffuse alveolar damage.

The definitions of acute exacerbations in chronic HP have been proposed as shown in **Box 1**.^{26,27} As in IPF, acute exacerbations predict poor outcome. The 2-year frequency of an acute exacerbation is 11.5%.²⁶

IMAGING FINDINGS: CHEST RADIOGRAPHY

On the chest radiograph, combined findings of transient diffuse ground-glass attenuation, airspace consolidation, micronodules, reticular shadows, and honeycombing are

Box 1 Definition of acute exacerbation of HP

1. Prior diagnosis of chronic HP
2. Worsening of dyspnea within 1 to 2 months
3. New radiographic opacities
4. Absence of apparent infection, heart disease, and/or other identifiable cause

prominent according to the clinical subforms of HP. In acute HP, diffuse ground-glass attenuation (GGA) and/or airspace consolidation, associated with some micronodules, may be seen. In subacute HP, micronodules, GGA, and reticular shadows are prominent. In chronic HP, reticular shadows and honeycombing are more predominant. In contrast with IPF, the changes are diffuse or may show upper zone predominance.

Mild enlargement of the mediastinal lymph nodes is occasionally found. Pleural involvement is rare. Clinicians should be aware that the chest radiograph may be normal in up to 30% of patients with HP.

IMAGING FINDINGS: HRCT

HRCT is useful in detecting HP and in separating the clinical subforms of HP. In acute HP, HRCT may be normal.²⁸ When abnormal, the characteristic findings on HRCT are patchy or diffuse GGA and/or centrilobular poorly defined small nodules; consolidation is rarely seen.^{29–34} Mosaic perfusion (air trapping) caused by concomitant bronchiolitis is also observed. This finding represents indirect signs of small airway obstruction. These small nodules are the common characteristics in not only acute but subacute or chronic HP (Fig. 1).

In subacute HP, patchy air-trapping areas on expiratory scans become more prominent, often in a lobular distribution.^{30,35} Because of the considerable overlap in subacute and chronic HP, the findings in chronic HP may be observed in subacute HP to varying degrees.

In chronic HP, the prominent findings on HRCT are the signs of lung fibrosis combined with GGA and centrilobular small nodules. The signs of lung fibrosis include interlobular septal thickening, lobar volume loss, linear-reticular opacities, traction bronchiectasis, and honeycombing (Fig. 2).^{29,36}

The reticulation is often distributed in the peribronchovascular area and lacks lower zone or subpleural predominance as in IPF. HRCT seems to be useful to distinguish IPF and NSIP from HP in many cases.^{36,37} In 1 study, the most characteristic findings in NSIP compared with chronic HP were the subpleural sparing, absence of lobular areas with GGA, and lack of honeycombing.³⁷ The most characteristic findings in IPF compared with chronic HP were the basal predominance of honeycombing, absence of relative subpleural sparing, and absence of centrilobular nodules. Honeycombing was seen in 64% of patients with chronic HP, which was as high a frequency as in IPF.³⁶

Additional emphysema can be seen in 20% of nonsmoking patients with chronic HP.^{32–34} Patients with chronic farmer's lung are more likely to develop emphysema than fibrosis.²²

Subacute and chronic HP sometimes show thin-walled cysts in areas of ground-glass attenuation, which mimic those observed in lymphocytic interstitial pneumonia.^{36,38}

PULMONARY FUNCTION TESTS

Although lung function may be normal in acute HP,²⁸ abnormal lung function is common in most patients with chronic HP. The most frequent functional abnormalities are a restrictive impairment and/or an impaired gas exchange (decreased diffusing capacity or increased alveolar/arterial oxygen gradient). Only few patients with farmer's lung show obstructive impairment resulting from emphysema.³⁹ However, these changes are not characteristic for chronic HP but are also found in any type of ILDs. Therefore, these abnormalities are not diagnostic for HP. Although hypoxemia is common in HP, patients with mild to moderate disease may lack this symptom and only present hypoxemia with exercise.

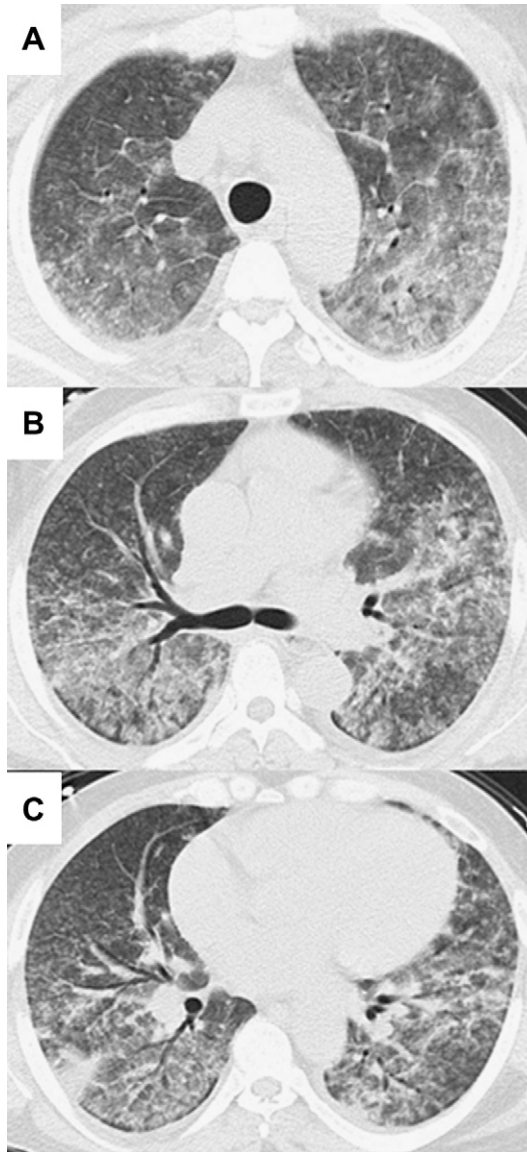


Fig. 1. HRCT of a patient with acute HP showing bilateral ground-glass densities with centrilobular micronodular accentuation and minor consolidation. (A) Upper lobes, (B) middle lobes, (C) lower lobes.

The functional impairment is not well correlated with the severity of radiological abnormalities. The importance of pulmonary function tests is to evaluate the severity of the physiologic impairment at diagnosis and during follow-up.²⁴

BAL AND INDUCED SPUTUM

BAL is a highly sensitive method to detect HP. An increase in the total cell count (usually more than 20 million in a total of 100 mL of BAL fluid) with a large increment

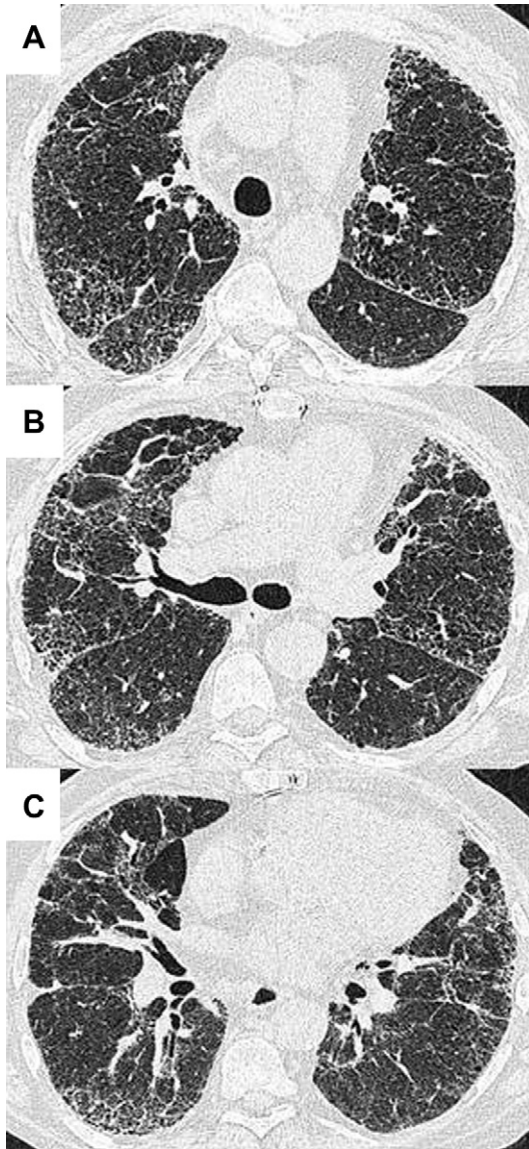


Fig. 2. HRCT of a patient with chronic HP showing bilateral reticular shadowing, traction bronchiectasis, and minor mosaic perfusion along with some micronodules. (A) Upper lobes, (B) middle lobes, (C) lower lobes.

of lymphocytes (usually more than 50%) is characteristic for HP, but not specific.⁴⁰ BAL lymphocytes show the highest count in HP of all ILDs. This increase is unusual in other differential diagnoses, including IPF.^{41,42} However, in patients with chronic HP or smokers, the increase in BAL lymphocytes may be less prominent.^{23,32,43} In contrast, even asymptomatic sensitized individuals may show increased lymphocytes in BAL fluid.^{44,45}

The evaluation of CD4+ and CD8+ T-cell subsets usually shows a relative predominance of CD8+ T cells resulting in a low CD4/CD8 ratio with mean values ranging between 0.5 and 1.0. However, the routine evaluation of the CD4/CD8 ratio is not recommended for clinical practice, because the various studies showed no consistent findings in CD4/CD8 ratio. The reasons for this discrepancy are unclear. The possible confounding factors may include the different disease manifestations (eg, the mean value of the CD4/CD8 ratio is higher in chronic HP than in subacute HP⁴⁶), the timing of BAL investigations, the type of inhaled antigen, the intensity of exposure, the smoking habit, and the clinical stage.^{24,42,46–48} CD4/CD8 ratios are increased within 24 hours after the last antigen exposure, and become lowest between 7 and 30 days.⁴⁹ Persistent BAL abnormalities during the follow-up may indicate incomplete antigen avoidance.

Small numbers of neutrophils, eosinophils, mast cells, and, more characteristically, plasma cells are also found in BAL fluid.^{32,49–52} The number of plasma cells in BAL fluid and immunoglobulin levels revealed a positive correlation, suggesting that the local production of immunoglobulins by plasma cells may play a pathogenetic role in susceptible individuals.⁵¹

Other morphologic features include signs of T-cell and macrophage activation.⁵³ Activated T cells show folded nuclei and/or broad cytoplasm, and have increased expression of counterligand CD28. Activated macrophages show foamy macrophages, and have increased expression of CD80/CD86.⁵⁴

The proteomic analysis of BAL fluids in HP seems to be useful for differentiating usual interstitial pneumonia (UIP) pattern from NSIP pattern.⁵⁵ Surfactant protein A, immunoglobulin heavy chain α , heat shock glycoprotein, haptoglobin β , and immunoglobulin J chain were increased in patients with UIP pattern, whereas glutathione S-transferase, vitamin D-binding protein, and β -actin were increased in patients with NSIP pattern.

In induced sputum from patients with HP, total cells and lymphocytes are also increased.⁵⁶ Differential cell counts showed that induced sputum and BAL reflected different compartments of inflammation.⁵⁶ A recent study showed that the CD4/CD8 ratio recovered from induced sputum is as useful as that recovered from BAL fluid.⁵⁷ Therefore, induced sputum may be complementary, but not an alternative, to BAL. The usefulness of induced sputum in the clinical practice or research use for HP is currently unclear.

LABORATORY TESTS

The presence of specific immunoglobulin (Ig) G antibodies (serum precipitins) to the exposed antigen is evidence of sensitization but not of disease. However, a positive test can be complementary for the diagnosis of HP and give the clinician useful additional information.⁵⁸

Approximately 10% of asymptomatic farmers and 40% of asymptomatic pigeon breeders show positive precipitating antibodies to the exposed antigens.^{59–61} Negative precipitating antibodies do not exclude the diagnosis of HP.^{62–64}

Various serologic/immunologic techniques including immune-electrophoresis enzyme immunoassay, fluoroenzyme immunoassay, peptide nucleic acid-fluorescence in situ hybridization, and DNA-fluorescence in situ hybridization assays were found to be useful for detecting HP antigens.^{65–67} Enzyme-linked immunosorbent assay is usually the preferred method. Increasing IgG antibody titers are correlated with the likelihood of HP, and decreasing titers reflect antigen avoidance.⁶⁸

A recent study enrolling a total of 122 patients with a suspected HP (including 31 cases of true HP) evaluated the diagnostic value of serum precipitins to mold antigens in HP, and showed that negative predictive values varied from 81% to 88% and

positive predictive values varied from 71% to 75%.⁵⁸ The selection of antigens to be tested needs to be determined based on the local predominant antigens.^{69,70}

In acute HP, the neutrophil fraction of the white blood cell count and the levels of C-reactive protein are increased. In chronic HP, polyclonal hypergammaglobulinemia is frequent. The rheumatoid factor may be positive in 50% of patients with pigeon HP.⁷¹

PROVOCATION TESTS

Inhalative provocation tests with the suspected antigen should only be performed in selected patients, because of the risk of a severe attack and the lack of standard procedure.²⁴ A natural exposure to the workplace or home seems to be a safer and more reasonable way to provoke symptoms.^{72,73}

Positive provocation findings typically include cough, dyspnea, fever, decrease in forced vital capacity and oxygen desaturation 8 to 12 hours after exposure. Because of the severity of the attack, patients should be monitored closely for at least 24 hours.

PATHOGENESIS

The pathogenesis of HP is complex, and the mechanisms involved are poorly understood. The presence of circulating precipitins to the relevant exposed antigens supported the concept that the disease is mediated by the deposition of antigen/antibody complexes within the alveolar walls (type III hypersensitivity).

However, several findings are not consistent with this hypothesis: (1) patients may develop disease but may lack serum precipitins, (2) histopathology does not show vasculitis or prominent neutrophil infiltration, and (3) in animal models passive serum transfer followed by aerosol exposure is not able to induce histologic changes of HP.⁶²

Histology of lymphocytic interstitial infiltrates with granuloma formation and signs of macrophage and lymphocyte activation in BAL may suggest a cell-mediated immune reaction (type IV hypersensitivity).⁵⁰

There is evidence for overproduction of Th1 cytokines⁷⁴ (interferon- γ , interleukin [IL]-12, and IL-18) along with tumor necrosis factor (TNF) receptors,⁷⁵ counterregulators of TNF, by alveolar macrophages from patients with HP.

Although HP is typically defined as Th1 disease, chronic HP evolving to fibrosis seems to be characterized by a switch to a Th2-biased immune response. The BAL fluid analyses from patients with chronic HP show overproduction of a Th2 chemokine family (CXC chemokine receptor [CXCR] 4, thymus and activation-regulated chemokine [TARC]/C-C motif ligand [CCL] 17), and downregulation of a Th1 chemokine family (CXCR3, interferon γ -induced protein [IP]-10, interferon- γ).^{46,76}

Although the mechanisms of HP have been partially clarified, it is still unclear why the disease develops only in a minority of exposed individuals. A 2-hit hypothesis suggested that the presence of an inducing factor (inhaled antigen) and a promoting factor (genetic susceptibility) may be essential for the development of HP. Several gene polymorphisms including TNF- α , transporter associated with antigen processing (TAP) genes, and the low-molecular-weight proteasome (LMP) 7 gene have been shown to be involved in the susceptibility HP.⁷⁷⁻⁸⁰ By contrast, polymorphisms in the tissue inhibitor of metalloproteinase (TIMP)-3 promoter gene may protect against the development of HP.^{81,82}

Toll-like receptors (TLRs) are expressed on immune cells and recognize various antigens. When specific TLRs are activated, many proinflammatory cytokines and mediators are released through an intracellular pathway (MyD88 pathway).⁸³ In experimental models of HP, the expression of TLR-9 and CD34 are essential for the development of a Th1 granulomatous inflammatory response.^{84,85}

Despite this progress, it is still not known why some patients show resolution of disease and others progress to fibrosis even without further antigen exposure.

PATHOLOGY

The difficulty in the interpretation of pathology results from the lack of a gold standard defining HP. Pathologic analyses in acute HP are rare. A retrospective study of selected cases of acute HP showed nonspecific diffuse pneumonitis and interstitial inflammation in a peribronchiolar pattern with mononuclear cell and neutrophil infiltration and fibrin deposition.¹ Intra-alveolar fibrin accumulation may be prominent in some selected cases with acute fibrinous and organizing pneumonia (AFOP).

Subacute HP is characterized by a lymphocytic, bronchiolocentric interstitial pneumonitis. The central regions of the secondary lobule are the predominant site to be involved.⁸⁶ It is independent of the presence or absence of further antigen exposure. Lymphocytes with fewer plasma cells and histiocytes are the main cells associated with inflammation. The granulomas are typically small, loose, poorly formed, and non-necrotizing, with the exception of hot-tub lung. Granulomatous changes may be absent in approximately 30% of patients with HP.⁸⁷ The staining with cathepsin K, a cysteine protease expressed in activated macrophages and epithelioid cells, may be useful for detecting microgranulomas in HP.⁸⁶ Cathepsin K staining is negative in patients with desquamative interstitial pneumonia (DIP) and respiratory bronchiolitis-ILD (RB-ILD), in which accumulation of alveolar macrophages is prominent. These findings suggest that cathepsin K may be a sensitive and specific marker to detect granulomas in chronic HP.

Chronic HP is characterized by progressive fibrosis, bronchiolitis obliterans, and architectural distortion in addition to the subacute changes. However, chronic HP may lack typical subacute changes.⁴³ The pathologic patterns may mimic UIP, NSIP, OP, or peribronchiolar interstitial fibrosis.^{9,43} In late chronic stages, the pathologic findings may become more similar to IPF/UIP.

The characteristic pathologic findings supporting HP includes bronchiolocentric inflammation, peribronchiolar fibrosis, bronchiolar epithelial hyperplasia, and the presence of granulomas or multinucleated giant cells.²⁵ Peribronchiolar metaplasia is frequently observed in HP, but is rare in IPF/UIP.⁸⁸ Peribronchiolar (centrilobular) fibrosis often extends to the perilobular areas, and forms the appearance of bridging fibrosis. This pathologic finding can distinguish chronic HP from IPF.⁸⁹

Although pathologic changes in HP are uniform in distribution, lung biopsy specimens sometimes show discordant findings.⁸⁸ This observation suggests that biopsy should be taken from at least 2 different lobes, as in IPF.

A recent study reported the coexistence of HP and pulmonary alveolar proteinosis (PAP).⁹⁰ Although all patients had typical HRCT findings of PAP to a varying degree, typical HRCT findings of HP were sometimes absent. The linkage between HP and PAP is still unclear.

DIAGNOSTIC CRITERIA

Several diagnostic criteria for HP have been recommended.^{91,92} However, none of these criteria has been validated. The diagnosis of HP relies on a high level of clinical suspicion; the recognition of antecedent antigen exposure; and a constellation of clinical, radiologic, laboratory, and pathologic findings.

A large prospective multicenter cohort study (116 patients with HP, 284 control subjects with other ILD) showed that the diagnosis of HP could be made with 6

significant predictors (**Box 2**). If all of the 6 predictors are present, the probability of having HP is 98%.⁶⁹ If none of the 6 predictors are present, the probability is 0%.⁶⁹

Careful history taking is mandatory. Clinicians should have specific expertise concerning the relevant antigens to HP. Important factors are hay feeding, bird keeping, feather duvet and pillows in the home, air conditioning or ventilators in the buildings, and formation of mold on room walls or in the cellars. Indirect contact with birds should also be asked for, such as visits to friends or relatives who keep birds in their homes. Improvement on vacation or during hospitalization may also be a clue to the diagnosis.

HRCT is a useful diagnostic test. Although it may be normal in some patients, the sensitivity is more than 95%, and the finding of a centrilobular micronodular ground-glass pattern and evidence of mosaic perfusion (trapped air) is characteristic of HP. The major differential diagnosis in this setting is respiratory bronchiolitis/ILD or *Pneumocystis carinii* infection.

The most sensitive diagnostic test is BAL. In the author's experience and based on literature review, a normal BAL widely excludes the diagnosis of HP. The characteristic finding is a lymphocytosis in the subacute and chronic forms. In asymptomatic sensitized individuals (subclinical alveolitis), BAL lymphocytosis is also apparent. BAL lymphocytosis greater than 30% is recommended as a discriminative factor of chronic HP showing UIP pattern on HRCT from IPF.⁴¹

BAL analyses have complementary information on HRCT. Lymphocytosis is characteristic for HP, a predominance of smoker's macrophages is characteristic for RB-ILD, and the presence of microorganisms is characteristic for *Pneumocystis carinii* pneumonia.

Pathologic evaluation of lung tissue is usually unnecessary for the diagnosis of HP. If needed, the preferred approach is surgical biopsy rather than transbronchial biopsy.

An important problem in the diagnosis of HP is that the relevant antigen cannot be identified in up to 20% to 30% of patients. In these patients the diagnosis must be suspected based on histopathology, BAL findings, and HRCT characteristics.⁹³

DIFFERENTIAL DIAGNOSIS

The differential diagnoses include the wide spectrum of ILD, mainly idiopathic interstitial pneumonias (IIPs) and sarcoidosis.⁶⁹ Frequent misdiagnosis is pneumonia in acute forms and chronic bronchitis in chronic forms with normal chest radiograph, which may occur in 20%. Chronic HP, especially the insidious form of bird fancier's lung, may closely mimic IPF or idiopathic fibrotic NSIP.⁹⁴

Clinicians should be aware that HP must be considered in all cases of ILD, and a detailed environmental exposure history is mandatory. **Fig. 3** shows the diagnostic algorithm for HP.

Box 2 Diagnosis of HP

1. Exposure to a known offending antigen
2. Positive precipitating antibodies
3. Recurrent episodes of symptoms
4. Inspiratory crackles
5. Symptoms 4 to 8 hours after exposure
6. Weight loss

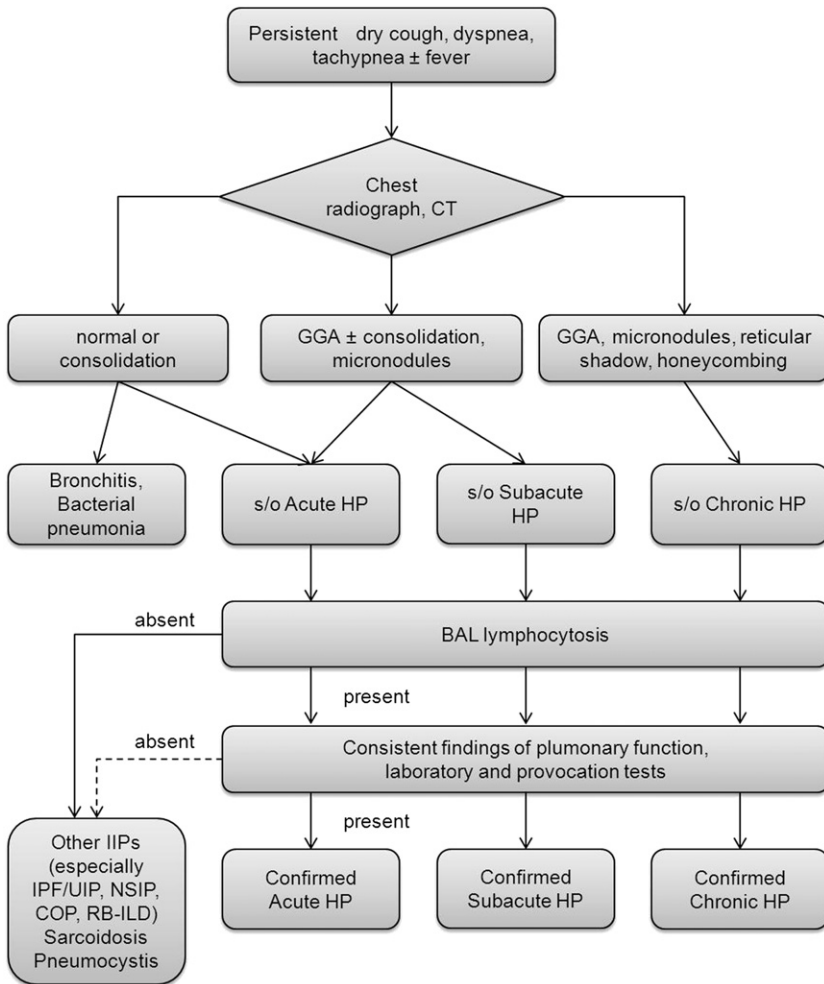


Fig. 3. Flow chart of diagnostic algorithm for HP. BAL, bronchoalveolar lavage; COP, cryptogenic organizing pneumonia; CT, computed tomography; NSIP, nonspecific interstitial pneumonia; RB-ILD, respiratory bronchiolitis-interstitial lung disease.

PITFALLS: EFFECT OF CIGARETTE SMOKING

HP is less frequent in smokers than in nonsmokers under the same exposure.²⁴ Cigarette smoking seems to protect against the development of HP. When exposed to high levels of antigens, smokers have lower levels of specific antibodies to the causative antigen compared with nonsmokers.

Although the mechanisms of the protective effect of smoking against HP are unclear, nicotine seems to be one of the key factors.⁹⁵ Nicotine inhibits macrophage activation, decreases lymphocyte proliferation, and impairs T-cell function.^{95,96}

Although HP develops more frequently in nonsmokers, when HP occurs in smokers, the patients may develop a chronic clinical course with more recurrent episodes and a significantly poorer survival compared with nonsmoker patients.⁹⁷

MANAGEMENT

Early diagnosis and antigen avoidance are key factors in the management of HP. Although complete avoidance of antigen exposure is difficult in some patients with HP, sustained antigen inhalation is associated with a poorer outcome.

Antigens may persist in rooms where birds have been kept for a long time. Feather pillows and blankets should be removed. Indirect and occasional exposure in the homes of friends or relatives where birds are kept should also be avoided. It is important to minimize microbial or avian antigen exposure by having a clean environment at home. The use of air-purifying respirators may be useful in some cases. Farmers should wear dust masks with filters, and ensure appropriate ventilation. Mechanization of the feeding process on farms and alterations in forced-air ventilatory systems may also be useful.

Corticosteroid therapy is usually recommended in patients who show functional impairment, although its long-term efficacy has not been evaluated in prospective clinical trials. Treatment continues until no further improvement in physiologic abnormalities is observed. An empiric therapy schedule may consist of 40 to 50 mg per day of prednisone for a month, followed by a gradual tapering during the next 2 to 3 months and a maintenance dose between 7.5 and 15 mg per day.

In chronic progressive HP, immunosuppressants may be added as corticosteroid sparing agents, as is done in other fibrotic ILDs.⁹⁴

Routine follow-up investigations should be more narrow immediately after diagnosis and during treatment (1–3 months is appropriate); later the interval can be extended to every 6 to 12 months. If the course is favorable, with complete remission after avoidance of further exposure and/or corticosteroid treatment, routine follow-up can be stopped after 2 to 3 years.

Inhaled steroids or pentoxifylline may be other options of treatment⁹⁸; however, their efficiency has not yet been validated.

In chronic progressive HP not responding to corticosteroid and/or immunosuppressant therapy, lung transplantation should be recommended.

PROGNOSIS

The prognosis of HP is variable and depends on the type, duration, and intensity of antigen exposure; the type of pathologic changes (UIP, NSIP, OP-like fibrosis, or emphysema); and possibly genetic background.⁹ The findings of fibrosis at lung biopsy or HRCT are associated with poor prognosis in patients with chronic HP and may serve as a useful prognostic indicator.⁹³

Patients with OP-like or cellular NSIP-like fibrosis have a more favorable outcome than those with fibrotic NSIP-like and UIP-like fibrosis.⁹⁴ The UIP-like and fibrotic NSIP-like fibrosis are associated with decreased survival.⁹⁹

Some patients may experience progression, despite avoiding exposure and undergoing treatment. There is no good explanation for the mechanism. Acute exacerbation of chronic HP is associated with a poor prognosis.²⁶ In a previous study enrolling 100 consecutive patients with chronic bird farmer's lung, 14 patients developed an acute exacerbation, and 12 of them died of this episode.²⁶

A previous surveillance in the United States showed that overall age-adjusted death rates in HP increased significantly ($P < .0001$) between 1980 and 2002, from 0.09 to 0.29 per million, although it is unclear what factors were associated with this increase.¹⁰⁰ By contrast, another surveillance in England and Wales showed that the mortality in HP was almost stable between 1968 and 2008, from 0.04 to 0.08 per million.¹⁰¹

In general, acute HP seems to have a favorable prognosis. After acute attacks, if correctly and timely diagnosed and treated, patients usually have a complete remission. If acute attacks occur frequently, such as in some patients with farmer's lung, the outcome may be the development of emphysema.

Patients with farmer's lung who experienced recurrent attacks tend to have emphysema more frequently and lower diffusing capacity than patients who experienced only a single attack.¹⁰²

Complications of lung cancer may affect the prognosis in HP. A recent retrospective review on 104 patients with chronic HP showed that the prevalence of lung cancer in chronic HP seems to be increased (10.6%), as seen in IPF.¹⁰³

Pulmonary hypertension occurs in approximately 20% of patients with chronic HP, and is associated with a greater risk of death (see **Box 2**).¹⁰⁴

SUMMARY

HP is a complex syndrome caused by repeated inhalation of environmental and occupational antigens. Clinical manifestations of HP may closely mimic other ILDs, including IPF or NSIP, and the disease onset is usually insidious; diagnosis of HP is therefore sometimes difficult. An appropriate removal of antigen exposure is essential for treatment of HP, otherwise the disease results in poor outcomes. Therefore, clinicians should be aware that HP must be considered in all cases of ILD, and should start the appropriate management as soon as possible.

REFERENCES

1. Costabel U, Bonella F, Guzman J. Chronic hypersensitivity pneumonitis. *Clin Chest Med* 2012;33:151–63.
2. Gruchow HW, Hoffmann RG, Marx JJ Jr, et al. Precipitating antibodies to farmer's lung antigens in a Wisconsin farming population. *Am Rev Respir Dis* 1981;124:411–5.
3. Terho EO, Heinonen OP, Lammi S, et al. Incidence of clinically confirmed farmer's lung in Finland and its relation to meteorological factors. *Eur J Respir Dis Suppl* 1987;152:47–56.
4. Depierre A, Dalphin JC, Pernet D, et al. Epidemiological study of farmer's lung in five districts of the French Doubs province. *Thorax* 1988;43:429–35.
5. Rodríguez de Castro F, Carrillo T, Castillo R, et al. Relationships between characteristics of exposure to pigeon antigens. Clinical manifestations and humoral immune response. *Chest* 1993;103:1059–63.
6. Hendrick DJ, Faux JA, Marshall R. Budgerigar-fancier's lung: the commonest variety of allergic alveolitis in Britain. *Br Med J* 1978;2:81–4.
7. Grech V, Vella C, Lenicker H. Pigeon breeder's lung in childhood: varied clinical picture at presentation. *Pediatr Pulmonol* 2000;30:145–8.
8. Ratjen F, Costabel U, Griesse M, et al. Bronchoalveolar lavage fluid findings in children with hypersensitivity pneumonitis. *Eur Respir J* 2003;21:144–8.
9. Lacasse Y, Girard M, Cormier Y. Recent advances in hypersensitivity pneumonitis. *Chest* 2012;142:208–17.
10. Koschel D, Stark W, Karmann F, et al. Extrinsic allergic alveolitis caused by misting fountains. *Respir Med* 2005;99:943–7.
11. Kämpfer P, Engelhart S, Rolke M, et al. Extrinsic allergic alveolitis (hypersensitivity pneumonitis) caused by *Sphingobacterium spiritivorum* from the water reservoir of a steam iron. *J Clin Microbiol* 2005;43:4908–10.

12. Morell F, Cruz MJ, Gómez FP, et al. Chaciner's lung - hypersensitivity pneumonitis due to dry sausage dust. *Scand J Work Environ Health* 2011;37:349–56.
13. Rouzaud P, Soulat JM, Trela C, et al. Symptoms and serum precipitins in workers exposed to dry sausage mould: consequences of exposure to sausage mould. *Int Arch Occup Environ Health* 2001;74:371–4.
14. Metzger F, Haccuria A, Reboux G, et al. Hypersensitivity pneumonitis due to molds in a saxophone player. *Chest* 2010;138:724–6.
15. Lodha S, Sharma OP. Hypersensitivity pneumonitis in a saxophone player. *Chest* 1988;93:1322.
16. Metersky ML, Bean SB, Meyer JD, et al. Trombone player's lung: a probable new cause of hypersensitivity pneumonitis. *Chest* 2010;138:754–6.
17. Leong KW, Ong S, Chee HL, et al. Hypersensitivity pneumonitis due to high-dose colistin aerosol therapy. *Int J Infect Dis* 2010;14:e1018–9.
18. Otera H, Tada K, Sakurai T, et al. Hypersensitivity pneumonitis associated with inhalation of catechin-rich green tea extracts. *Respiration* 2011;82:388–92.
19. Scherpereel A, Tillie-Leblond I, Pommier de Santi P, et al. Exposure to methyl methacrylate and hypersensitivity pneumonitis in dental technicians. *Allergy* 2004;59:890–2.
20. Koschel D, Lützkendorf L, Wiedemann B, et al. Antigen-specific IgG antibodies in feather duvet lung. *Eur J Clin Invest* 2010;40:797–802.
21. Koschel D, Wittstruck H, Renck T, et al. Presenting features of feather duvet lung. *Int Arch Allergy Immunol* 2010;152:264–70.
22. Malinen AP, Erkinjuntti-Pekkanen RA, Partanen PL, et al. Long-term sequelae of farmer's lung disease in HRCT: a 14-year follow-up study of 88 patients and 83 matched control farmers. *Eur Radiol* 2003;13:2212–21.
23. Ohtani Y, Saiki S, Sumi Y, et al. Clinical features of recurrent and insidious chronic bird fancier's lung. *Ann Allergy Asthma Immunol* 2003;90:604–10.
24. Selman M, Pardo A, King TE Jr. Hypersensitivity pneumonitis: insights in diagnosis and pathobiology. *Am J Respir Crit Care Med* 2012;186(4):314–24.
25. Churg A, Muller NL, Flint J, et al. Chronic hypersensitivity pneumonitis. *Am J Surg Pathol* 2006;30:201–8.
26. Miyazaki Y, Tateishi T, Akashi T, et al. Clinical predictors and histologic appearance of acute exacerbations in chronic hypersensitivity pneumonitis. *Chest* 2008;134:1265–70.
27. Olson AL, Huie TJ, Groshong SD, et al. Acute exacerbations of fibrotic hypersensitivity pneumonitis: a case series. *Chest* 2008;134:844–50.
28. Lynch DA, Rose CS, Way D, et al. Hypersensitivity pneumonitis: sensitivity of high-resolution CT in a population-based study. *AJR Am J Roentgenol* 1992;159:469–72.
29. Tateishi T, Ohtani Y, Takemura T, et al. Serial high-resolution computed tomography findings of acute and chronic hypersensitivity pneumonitis induced by avian antigen. *J Comput Assist Tomogr* 2011;35:272–9.
30. Patel RA, Sellami D, Gotway MB, et al. Hypersensitivity pneumonitis: patterns on high-resolution CT. *J Comput Assist Tomogr* 2000;24:965–70.
31. Adler BD, Padley SP, Müller NL, et al. Chronic hypersensitivity pneumonitis: high-resolution CT and radiographic features in 16 patients. *Radiology* 1992;185:91–5.
32. Remy-Jardin M, Remy J, Wallaert B, et al. Subacute and chronic bird breeder hypersensitivity pneumonitis: sequential evaluation with CT and correlation with lung function tests and bronchoalveolar lavage. *Radiology* 1993;189:111–8.

33. Erkinjuntti-Pekkanen R, Rytönen H, Kokkarinen JI, et al. Long-term risk of emphysema in patients with farmer's lung and matched control farmers. *Am J Respir Crit Care Med* 1998;158:662–5.
34. Cormier Y, Brown M, Worthy S, et al. High-resolution computed tomographic characteristics in acute farmer's lung and in its follow-up. *Eur Respir J* 2000;16:56–60.
35. Hansell DM, Wells AU, Padley SP, et al. Hypersensitivity pneumonitis: correlation of individual CT patterns with functional abnormalities. *Radiology* 1996;199:123–8.
36. Silva CI, Müller NL, Lynch DA, et al. Chronic hypersensitivity pneumonitis: differentiation from idiopathic pulmonary fibrosis and nonspecific interstitial pneumonia by using thin-section CT. *Radiology* 2008;246:288–97.
37. Lynch DA, Newell JD, Logan PM, et al. Can CT distinguish hypersensitivity pneumonitis from idiopathic pulmonary fibrosis? *AJR Am J Roentgenol* 1995;165:807–11.
38. Franquet T, Hansell DM, Senbanjo T, et al. Lung cysts in subacute hypersensitivity pneumonitis. *J Comput Assist Tomogr* 2003;27:475–8.
39. Lalancette M, Carrier G, Laviolette M, et al. Farmer's lung. Long-term outcome and lack of predictive value of bronchoalveolar lavage fibrosing factors. *Am Rev Respir Dis* 1993;148:216–21.
40. Semenzato G, Bjermer L, Costabel U, et al. Clinical guidelines and indications for bronchoalveolar lavage (BAL): extrinsic allergic alveolitis. *Eur Respir J* 1990;3:945–6.
41. Ohshimo S, Bonella F, Cui A, et al. Significance of bronchoalveolar lavage for the diagnosis of idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med* 2009;179:1043–7.
42. Meyer KC, Raghu G, Baughman RP, et al. An official American Thoracic Society clinical practice guideline: the clinical utility of bronchoalveolar lavage cellular analysis in interstitial lung disease. *Am J Respir Crit Care Med* 2012;185:1004–14.
43. Gaxiola M, Buendía-Roldán I, Mejía M, et al. Morphologic diversity of chronic pigeon breeder's disease: clinical features and survival. *Respir Med* 2011;105:608–14.
44. Cormier Y, Bélanger J, Laviolette M. Persistent bronchoalveolar lymphocytosis in asymptomatic farmers. *Am Rev Respir Dis* 1986;133:843–7.
45. Cormier Y, Létourneau L, Racine G. Significance of precipitins and asymptomatic lymphocytic alveolitis: a 20-yr follow-up. *Eur Respir J* 2004;23:523–5.
46. Barrera L, Mendoza F, Zuñiga J, et al. Functional diversity of T-cell subpopulations in subacute and chronic hypersensitivity pneumonitis. *Am J Respir Crit Care Med* 2008;177:44–55.
47. Ando M, Konishi K, Yoneda R, et al. Difference in the phenotypes of bronchoalveolar lavage lymphocytes in patients with summer-type hypersensitivity pneumonitis, farmer's lung, ventilation pneumonitis, and bird fancier's lung: report of a nationwide epidemiologic study in Japan. *J Allergy Clin Immunol* 1991;87:1002–9.
48. Wahlström J, Berlin M, Lundgren R, et al. Lung and blood T-cell receptor repertoire in extrinsic allergic alveolitis. *Eur Respir J* 1997;10:772–9.
49. Drent M, van Velzen-Blad H, Diamant M, et al. Bronchoalveolar lavage in extrinsic allergic alveolitis: effect of time elapsed since antigen exposure. *Eur Respir J* 1993;6:1276–81.
50. Costabel U. The alveolitis of hypersensitivity pneumonitis. *Eur Respir J* 1988;1:5–9.
51. Drent M, Wagenaar S, van Velzen-Blad H, et al. Relationship between plasma cell levels and profile of bronchoalveolar lavage fluid in patients with subacute extrinsic allergic alveolitis. *Thorax* 1993;48:835–9.

52. Groot Kormelink T, Pardo A, Knipping K, et al. Immunoglobulin free light chains are increased in hypersensitivity pneumonitis and idiopathic pulmonary fibrosis. *PLoS One* 2011;6:e25392.
53. Costabel U, Bross KJ, Rühle KH, et al. Ia-like antigens on T-cells and their subpopulations in pulmonary sarcoidosis and in hypersensitivity pneumonitis. Analysis of bronchoalveolar and blood lymphocytes. *Am Rev Respir Dis* 1985; 131:337–42.
54. Blatman KH, Grammer LC. Chapter 19: hypersensitivity pneumonitis. *Allergy Asthma Proc* 2012;33(Suppl 1):64–6.
55. Okamoto T, Miyazaki Y, Shirahama R, et al. Proteome analysis of bronchoalveolar lavage fluid in chronic hypersensitivity pneumonitis. *Allergol Int* 2012;61:83–92.
56. D'Ippolito R, Chetta A, Foresi A, et al. Induced sputum and bronchoalveolar lavage from patients with hypersensitivity pneumonitis. *Respir Med* 2004;98: 977–83.
57. Economidou F, Samara KD, Antoniou KM, et al. Induced sputum in interstitial lung diseases: novel insights in the diagnosis, evaluation and research. *Respiration* 2009;77:351–8.
58. Fenoglio CM, Reboux G, Sudre B, et al. Diagnostic value of serum precipitins to mould antigens in active hypersensitivity pneumonitis. *Eur Respir J* 2007;29: 706–12.
59. Cormier Y, Bélanger J, Durand P. Factors influencing the development of serum precipitins to farmer's lung antigen in Quebec dairy farmers. *Thorax* 1985;40: 138–42.
60. Fink JN. Epidemiologic aspects of hypersensitivity pneumonitis. *Monogr Allergy* 1987;21:59–69.
61. Dalphin JC, Toson B, Monnet E, et al. Farmer's lung precipitins in Doubs (a department of France): prevalence and diagnostic value. *Allergy* 1994;49: 744–50.
62. Sennekamp J, Niese D, Stroehmann I, et al. Pigeon breeders' lung lacking detectable antibodies. *Clin Allergy* 1978;8:305–10.
63. Cormier Y, Bélanger J. The fluctuant nature of precipitating antibodies in dairy farmers. *Thorax* 1989;44:469–73.
64. Erkinjuntti-Pekkanen R, Reiman M, Kokkarinen JI, et al. IgG antibodies, chronic bronchitis, and pulmonary function values in farmer's lung patients and matched controls. *Allergy* 1999;54:1181–7.
65. Rodrigo MJ, Postigo I, Wangenstein O, et al. A new application of Streptavidin ImmunoCAP for measuring IgG antibodies against non-available commercial antigens. *Clin Chim Acta* 2010;411:1675–8.
66. Reboux G, Piarroux R, Roussel S, et al. Assessment of four serological techniques in the immunological diagnosis of farmers' lung disease. *J Med Microbiol* 2007;56:1317–21.
67. Selvaraju SB, Kapoor R, Yadav JS. Peptide nucleic acid-fluorescence in situ hybridization (PNA-FISH) assay for specific detection of mycobacterium immunogenum and DNA-FISH assay for analysis of pseudomonads in metalworking fluids and sputum. *Mol Cell Probes* 2008;22:273–80.
68. McSharry C, Dye GM, Ismail T, et al. Quantifying serum antibody in bird fanciers' hypersensitivity pneumonitis. *BMC Pulm Med* 2006;6:16.
69. Lacasse Y, Selman M, Costabel U, et al. Clinical diagnosis of hypersensitivity pneumonitis. *Am J Respir Crit Care Med* 2003;168:952–8.
70. Ojanen T. Class specific antibodies in serodiagnosis of farmer's lung. *Br J Ind Med* 1992;49:332–6.

71. Aguilar León DE, Novelo Retana V, Martínez-Cordero E. Anti-avian antibodies and rheumatoid factor in pigeon hypersensitivity pneumonitis. *Clin Exp Allergy* 2003;33:226–32.
72. Ramírez-Venegas A, Sansores RH, Pérez-Padilla R, et al. Utility of a provocation test for diagnosis of chronic pigeon breeder's disease. *Am J Respir Crit Care Med* 1998;158:862–9.
73. Ohtani Y, Kojima K, Sumi Y, et al. Inhalation provocation tests in chronic bird fancier's lung. *Chest* 2000;118:1382–9.
74. Yamasaki H, Ando M, Brazer W, et al. Polarized type 1 cytokine profile in bronchoalveolar lavage T cells of patients with hypersensitivity pneumonitis. *J Immunol* 1999;163:3516–23.
75. Dai H, Guzman J, Chen B, et al. Production of soluble tumor necrosis factor receptors and tumor necrosis factor- α by alveolar macrophages in sarcoidosis and extrinsic allergic alveolitis. *Chest* 2005;127:251–6.
76. Kishi M, Miyazaki Y, Jinta T, et al. Pathogenesis of cBFL in common with IPF? Correlation of IP-10/TARC ratio with histological patterns. *Thorax* 2008;63:810–6.
77. Camarena A, Juarez A, Mejia M, et al. Major histocompatibility complex and tumor necrosis factor- α polymorphisms in pigeon breeder's disease. *Am J Respir Crit Care Med* 2001;163:1528–33.
78. Schaaf BM, Seitzer U, Pravica V, et al. Tumor necrosis factor- α -308 promoter gene polymorphism and increased tumor necrosis factor serum bioactivity in farmer's lung patients. *Am J Respir Crit Care Med* 2001;163:379–82.
79. Aquino-Galvez A, Camarena A, Montano M, et al. Transporter associated with antigen processing (TAP) 1 gene polymorphisms in patients with hypersensitivity pneumonitis. *Exp Mol Pathol* 2008;84:173–7.
80. Camarena A, Aquino-Galvez A, Falfan-Valencia R, et al. PSMB8 (LMP7) but not PSMB9 (LMP2) gene polymorphisms are associated to pigeon breeder's hypersensitivity pneumonitis. *Respir Med* 2010;104:889–94.
81. Hill MR, Briggs L, Montano MM, et al. Promoter variants in tissue inhibitor of metalloproteinase-3 (TIMP-3) protect against susceptibility in pigeon breeders' disease. *Thorax* 2004;59:586–90.
82. Janssen R, Kruit A, Grutters JC, et al. TIMP-3 promoter gene polymorphisms in BFL. *Thorax* 2005;60:974.
83. Nance SC, Yi AK, Re FC, et al. MyD88 is necessary for neutrophil recruitment in hypersensitivity pneumonitis. *J Leukoc Biol* 2008;83:1207–17.
84. Bhan U, Newstead MJ, Zeng X, et al. *Stachybotrys chartarum*-induced hypersensitivity pneumonitis is TLR9 dependent. *Am J Pathol* 2011;179:2779–87.
85. Blanchet MR, Bennett JL, Gold MJ, et al. CD34 is required for dendritic cell trafficking and pathology in murine hypersensitivity pneumonitis. *Am J Respir Crit Care Med* 2011;184:687–98.
86. Reghellin D, Poletti V, Tomassett S, et al. Cathepsin-K is a sensitive immunohistochemical marker for detection of micro-granulomas in hypersensitivity pneumonitis. *Sarcoidosis Vasc Diffuse Lung Dis* 2010;27:57–63.
87. Myers JL. Hypersensitivity pneumonia: the role of lung biopsy in diagnosis and management. *Mod Pathol* 2012;25:S58–67.
88. Trahan S, Hanak V, Ryu JH, et al. Role of surgical lung biopsy in separating chronic hypersensitivity pneumonia from usual interstitial pneumonia/idiopathic pulmonary fibrosis: analysis of 31 biopsies from 15 patients. *Chest* 2008;134:126–32.
89. Akashi T, Takemura T, Ando N, et al. Histopathologic analysis of sixteen autopsy cases of chronic hypersensitivity pneumonitis and comparison with idiopathic

- pulmonary fibrosis/usual interstitial pneumonia. *Am J Clin Pathol* 2009;131:405–15.
90. Verma H, Nicholson AG, Kerr KM, et al. Alveolar proteinosis with hypersensitivity pneumonitis: a new clinical phenotype. *Respirology* 2010;15:1197–202.
 91. Terho EO. Diagnostic criteria for farmer's lung disease. *Am J Ind Med* 1986;10:329.
 92. Schuyler M, Cormier Y. The diagnosis of hypersensitivity pneumonitis. *Chest* 1997;111:534–6.
 93. Hanak V, Golbin JM, Hartman TE, et al. High-resolution CT findings of parenchymal fibrosis correlate with prognosis in hypersensitivity pneumonitis. *Chest* 2008;134:133–8.
 94. Ohtani Y, Saiki S, Kitaichi M, et al. Chronic bird fancier's lung: histopathological and clinical correlation. An application of the 2002 ATS/ERS consensus classification of the idiopathic interstitial pneumonias. *Thorax* 2005;60:665–71.
 95. Blanchet MR, Israël-Assayag E, Cormier Y. Inhibitory effect of nicotine on experimental hypersensitivity pneumonitis in vivo and in vitro. *Am J Respir Crit Care Med* 2004;169:903–9.
 96. Nizri E, Irony-Tur-Sinai M, Lory O, et al. Activation of the cholinergic anti-inflammatory system by nicotine attenuates neuroinflammation via suppression of Th1 and Th17 responses. *J Immunol* 2009;183:6681–8.
 97. Ohtsuka Y, Munakata M, Tanimura K, et al. Smoking promotes insidious and chronic farmer's lung disease, and deteriorates the clinical outcome. *Intern Med* 1995;34:966–71.
 98. Tanaka H, Tsunematsu K, Nakamura N, et al. Successful treatment of hypersensitivity pneumonitis caused by *Grifola frondosa* (Maitake) mushroom using a HFA-BDP extra-fine aerosol. *Intern Med* 2004;43:737–40.
 99. Vourlekis JS, Schwarz MI, Cherniack RM, et al. The effect of pulmonary fibrosis on survival in patients with hypersensitivity pneumonitis. *Am J Med* 2004;116:662–8.
 100. Bang KM, Weissman DN, Pinheiro GA, et al. Twenty-three years of hypersensitivity pneumonitis mortality surveillance in the United States. *Am J Ind Med* 2006;49:997–1004.
 101. Hanley A, Hubbard RB, Navaratnam V. Mortality trends in asbestosis, extrinsic allergic alveolitis and sarcoidosis in England and Wales. *Respir Med* 2011;105:1373–9.
 102. Erkinjuntti-Pekkanen R, Kokkarinen JI, Tukiainen HO, et al. Long-term outcome of pulmonary function in farmer's lung: a 14 year follow-up with matched controls. *Eur Respir J* 1997;10:2046–50.
 103. Kuramochi J, Inase N, Miyazaki Y, et al. Lung cancer in chronic hypersensitivity pneumonitis. *Respiration* 2011;82:263–7.
 104. Koschel DS, Cardoso C, Wiedemann B, et al. Pulmonary hypertension in chronic hypersensitivity pneumonitis. *Lung* 2012;190(3):295–302.